

Review

# Plants-occurring Anthraquinones as Photosensitive Compounds for Photodynamic Method: A Critical Overview

Vanya Mantareva<sup>1,\*</sup>, Diana Braikova<sup>1</sup>, Irina Lazarova<sup>2</sup>, Tsanislava Genova<sup>3</sup><sup>1</sup>Institute of Organic Chemistry with Centre of Phytochemistry, Bulgarian Academy of Sciences, 1113 Sofia, Bulgaria<sup>2</sup>Faculty of Pharmacy, Medical University – Sofia, Dunay, 1000 Sofia, Bulgaria<sup>3</sup>Institute of Electronics, Bulgarian Academy of Sciences, 1784 Sofia, Bulgaria\*Correspondence: [mantareva@yahoo.com](mailto:mantareva@yahoo.com); [Vanya.Mantareva@orgchm.bas.bg](mailto:Vanya.Mantareva@orgchm.bas.bg) (Vanya Mantareva)

Academic Editor: Jen-Tsung Chen

Submitted: 29 November 2023 Revised: 19 February 2024 Accepted: 8 March 2024 Published: 28 April 2024

## Abstract

The review focuses on the recent knowledge on natural anthraquinones (AQs) of plant origin and their potential for application in an exclusive medicinal curative and palliative method named photodynamic therapy (PDT). Green approach to PDT is associated with photosensitizers (PS) from plants or other natural sources and excitation light in visible spectrum. The investigations of plants are of high research interests due to their unique health supportive properties as herbs and the high percentage availability to obtain compounds with medical value. Up-to-date many naturally occurring compounds with therapeutic properties are known and are still under investigations. Some natural quinones have already been evaluated and clinically approved as anti-tumor agents. Recent scientific interests are beyond their common medical applications but also in directions to their photo-properties as natural PSs. The study presents a systematic searches on the latest knowledge on AQ derivatives that are isolated from the higher plants as photosensitizers for PDT applications. The natural quinones have been recognized with functions of natural dyes since the ancient times. Lately, AQs have been explored due to their biological activity including the photosensitive properties useful for PDT especially towards medical problems with no other alternatives. The existing literature' overview suggests that natural AQs possess characteristics of valuable PSs for PDT. This method is based on an application of a photoactive compound and light arrangement in oxygen media, such that the harmful general cytotoxicity could be avoided. Moreover, the common anticancer and antimicrobial drug resistance has been evaluated with very low occurrence after PDT. Natural AQs have been focused the scientific efforts to further developments because of the high range of natural sources, desirable biocompatibility, low toxicity, minimal side effects and low accident of drug resistance, together with their good photosensitivity and therapeutic capacity. Among the known AQs, only hypericin has been studied in anticancer clinical PDT. Currently, the natural PSs are under intensive research for the future PDT applications for diseases without alternative effective treatments.

**Keywords:** anthraquinones; natural photosensitizers; plants; herbs; reactive oxygen species; photodynamic therapy (PDT); biomedical applications

## 1. Introduction

The naturally occurring compounds including anthraquinones (AQs) have been under intensive investigations because of the fast development of drug resistance towards conventional chemo-drugs and routine therapeutics [1]. Moreover, there is a general tendency of back to the nature as a main source of low toxicity substances with comparable biological activity as the synthetic analogues. Worldwide, ~80% plants are used to treat diseases in everyday life [2]. As a result of the recognition of the numerous traditional routines in healthcare systems, these medicinal plants and herbs have increasingly been in free usages. Most of the natural AQ derivatives that have been isolated from plants are introduced to be photosensitive compounds in the ultraviolet (UV)-visible spectrum [3]. Among the natural photosensitizers (PS) only few of them such as chlorophyll, curcumin, pheophorbide A, hypocreline and AQs aloe-emodin, and hypericin have been studied for photodynamic therapy (PDT) method [4]. Nonetheless that

most of the natural occurring-PSs have been evaluated with properties apart of being ideal PS, their research and development as photosensitive compounds for PDT are still under research interests because of other advantages of their curative actions [5]. A better solution to the main problem with harsh dark and phototoxicity of synthetic PSs are the photosensitive compounds of natural origin.

Currently, only synthetic PDT drugs are in clinical practice [6]. The properties of the known AQs such as low water-solubility, insufficient selectivity, long lasting skin phototoxicity are leading to many complications and undesirable side effects including genotoxicity and mutagenicity. These drugs are under further improvement through different structural modifications. It is worth to refer to the nature for PDT drugs starting with the known as well as the novel natural photoactive compounds. Among the natural AQs, only hypericin (HYP) found in the plant species of the genus *Hypericum* L., especially *Hypericum perforatum* L. has already been clinically investi-



gated in cancer PDT [7]. Chemically HYP is a polymeric 1,8-dihydroxyanthraquinone (4,5,7,4',5',7'-hexahydroxy-2,2-dimethylnaphthodi anthrone) with a restriction of absorbance which is in UV and short visible range (<590 nm), situated out of the “optical PDT window” (630–850 nm) [8]. A limitation of natural PSs which includes the spectra of absorption within the sun light spectrum might lead to undesirable phototoxicity. Another one such as a minimal or lack of selectivity can provoke the harmful phototoxicity on normal cells as well together with skin phototoxicity [9].

One of the largest groups of natural quinones that can be found in plants, but also in fungi, insects and lichens are 1,8-dihydroxyanthraquinones [10]. Initially, AQs are discovered as natural colorants with variety of colors that make them usable as dyes and pigments in industrial coloration [11]. Their polycyclic planar, rigid aromatic structure contains 14 carbons in the fused benzene rings which is tricyclic anthracene with two carbonyl groups including two benzene rings. The largest class of quinones are oxidized forms of anthracene derivatives which include 1,4- and 9,10-anthraquinones creating dioxanthracene with characteristics of natural pigments.

The most studied plants containing AQs are part of mainly three families (81%): Fabaceae, Rubiaceae and Polygonaceae [12]. Additional families that contain AQ-derivatives are Asphodelaceae, Rhamnaceae, Hypericaceae, Scrophulariaceae, Bignoniaceae, Pedaliaceae, Verbenaceae and Valerianaceae. However, in higher amount AQs can be found in Fabaceae (*Cassia* spp. and *Senna* spp.), Rubiaceae (*Morinda* spp. and *Rubia* spp.), Polygonaceae (*Rheum* spp. and *Rumex* spp.), Rhamnaceae (*Rhamnus* spp. and *Berberis* spp.) and Asphodelaceae (*Aloe* spp. and *Asphodeline* spp.). Among the most identified AQ-derivatives are aloe emodin, chrysophanol, emodin, physcion, rhein, rubiadin and soranjidiol, and damnacanthal (**Supplementary Table 1**).

Recent literature reviews suggested that the plants' occurring AQs possess a broad spectrum of medical actions, including antimicrobial, antiviral, cathartic, anticancer, diuretic, vasorelaxant, phytoestrogen, autoimmune disorders and many other clinical applications [13,14]. The wavelength of HYP (~590 nm) used for photo-experiments is considered as limitation for PDT due to the lesser light penetration in disordered cells [15]. This determines the development of the synthetic AQs [16]. PDT procedure includes a multi-factorial step that generally lead to production of toxic oxygen species (Fig. 1). The photodynamic compounds aim to produce different reactive oxygen species (ROS) by two mechanisms including oxygen radicals and anions (Type I) and singlet oxygen (Type II). The both are initiated by photon energy and next the electron/ proton or an energy transfer reaction due to formation of excited-state PSs. The produced reactive species further affect the pathologic target cells by several mechanisms [17]. There are limited numbers of studies on natural AQs, which are re-

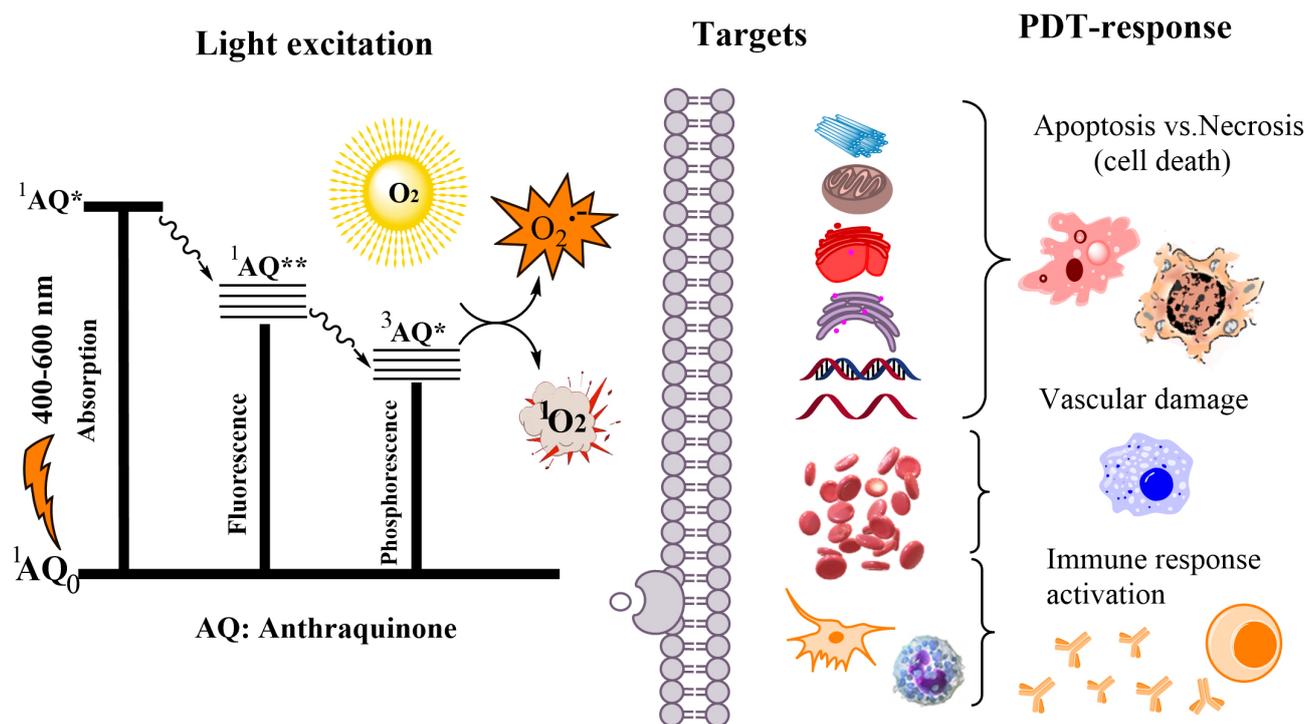
lated to possible mechanisms of PDT action [18–20]. The PDT response might lead to apoptosis or necrosis, degeneration of the tumor vasculature, stimulation of anti-tumor immune response, and inflammatory reactions in the treated lesion (Fig. 1).

The plant-occurring AQs are among the natural producers of anthracycline drugs which are observed as the most effective antineoplastic agents with planar tetracyclic molecules based on AQ core linked to a sugar moiety and phenolic groups [21]. The first clinically accepted compounds in cancer chemotherapy were daunorubicin and doxorubicin (DOX) with DNA binding by intercalation and the trapping of DNA Topoisomerase II complexes [22]. As a DNA intercalator, DOX forms hydrogen-bonds with guanine and insert into the DNA strand at sites with adjacent guanine-cytosine (GC) base pairs. This process tends to cause DNA damage and induce cell death. A second, Topoisomerase II poisoning by DOX interrupts DNA replication and transcription, resulting in apoptosis mediated cell death. The oxidation of DOX consequences in radical formation and then, the superoxide and H<sub>2</sub>O<sub>2</sub> production that leads to raised oxidative stress, which may lead to DNA damage [23]. The treatment also outcomes in an increased cellular ceramide level, activating growth arrest, apoptosis, and senescence. A catalytic action was observed as a result of undergoing cyclic one electron oxidation-reduction reactions. The produced ROS are capable of induction of oxidative stress causing membrane lyses and cellular damage, which consequently results in cell death [24,25].

This review summarizes on the recent studies of natural AQs originated from higher plants and the main plants' families with plant-originated AQs which are already studied as therapeutics. This is an attempt for a critical overview on the AQs as photosensitizers and their phototoxicity and photodynamic action. Additionally, we made a discussion on the chemical structure, and the correlation between the photosensitizing properties and biological PDT functions of the most investigated plants extracts and the isolated AQs. The limited AQ-PDT studies on different pathological targets (tumors, viruses, pathogenic bacteria, fungi and others) are described and discussed in respect to the potential usage of AQs in clinical PDT practice for an alternative curative therapy due to resistance and in emergency cases.

## 2. Materials and Methods

The article is based on the systematic overview on the research papers and reviews published in the last years considering the research field of our interest. The literature sources taken from the PubMed, Scholar Google, Web of Science and Scopus databases are issued during last ten years (till end of 2023) but not limited to this time for some themes. **Supplementary Material** summarizes in a Table the data about AQs with their structures and the plants of their origin during the first appearance in the literature until the time of manuscript preparation. The searches were



**Fig. 1. Mechanisms of anthraquinone (AQ) photo-excitation and the possible pathways of the photodynamic therapy (PDT) action and the targets.** \*: singlet or triplet excited state of AQ; \*\*: lowest energy excited state of AQ.

made using the keywords “anthraquinones”, “plants and herbs”, “photosensitizers” and “photosensitization”, “photodynamic therapy”, “phototoxicity”. The searches were limited to the most popular AQs isolated from higher plants which are already in usage as medicinal herbs with focus on the photo-properties and PDT studies.

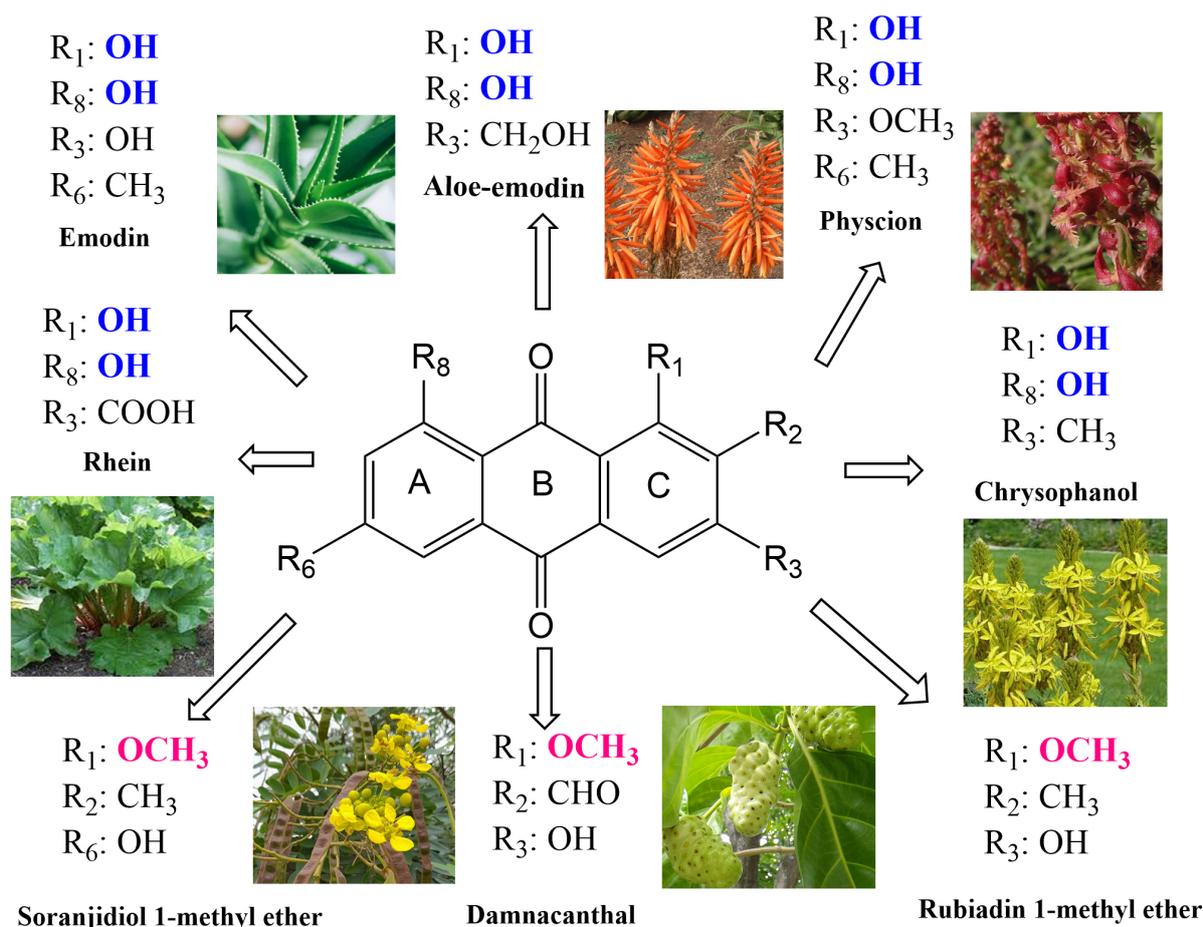
### 3. Plants-occurring Anthraquinone Photosensitizers

A great number of anthraquinones (AQs) have been isolated from higher plant species and subsequently characterized. Those higher plant species are spreading mainly in angiosperm of limited number of families [26]. In plants, fungi, and lichens, there are around 700 representatives species of this group. Compounds with origin from flowering plants which contain AQs as was lately described are approximately 200 structures [27]. Different quantities of AQs have been found in various parts of plant tissues namely flowers, roots, stems, rhizomes, bark, fruits and leaves like rhizomes. In those plants, where the biosynthesis goes by the shikimate-mevalonate pathway, the AQs predominate as aglycones [28,29]. Much more derivatives of AQs (~700) are described as useful dyes and pigments, making them the most significant group of naturally occurring colorants [30]. Natural AQs are well-studied secondary metabolites, products of biosynthesis that occurs via two major metabolic pathways, namely polyketide pathway (acetate-malonate pathway) and the shikimate pathway. The main difference between the both

biosynthetic pathways is that one produces 1,8-dihydroxy anthraquinones (acetate-malonate pathway) and the other (shikimate) pathway never generates substitution at position 8 (Fig. 2). The cathartic activity of some AQs is related to the 1,8-diOH substitution [26,30]. Anthraquinones from *Rubia* spp. are produced through the chorismite/o-succinylbenzoic acid or shikimate pathways [31], while the emodin type AQs (among them are the most effective photoactive compounds) are predominantly synthesized through the polyketide pathway [32].

#### 3.1 Fabaceae (*Cassia*, *Senna*)

The plants of the genus *Cassia* L. are an important source of naturally occurring AQs mainly used as bioactive compounds with antifungal, antibacterial, antiviral, insecticidal and other biological activity [33,34]. A large number of AQ structures were isolated from various parts of the *Cassia* spp. [35]. The numerous derivatives of AQs are characterized in these plants which are widely used in traditional folk medicines as therapeutics in antimalarial, ulcer healing, anti-diabetic, hepatoprotective, nephroprotective, antitumor and also in treatment of skin infection from the population over the tropical and subtropical regions [36–38]. Firstly, the plant extracts have been progressively used not only for cosmetics, foods and dyes but also for treatment of different medical problems and acute diseases due to their wide therapeutic and pharmacological properties [39,40]. Many *Cassia* spp. exist worldwide that are mostly used in herbal medicine [41]. These particular



**Fig. 2. Anthraquinones from higher plants with properties for PDT.** Their common substitution groups are in color and close to the plants.

family of plants is known for their laxative actions. *Cassia fistula* L. has the capacity of curative important and/or nutritionally valuable source. Leaves of *Cassia fistula* mainly contain AQ-derivatives together with other group of compounds (quinones, oxalic acids and tannins). The phytochemical investigation reveals that this specie contains a mixture of chrysophanol, physcion, rhein and other non-AQ compounds [42]. The reduced toxicity and minimal side effects of natural AQs observed in *Cassia* plants feature their potential as therapeutics. A recent report of Oliveira *et al.* [43] showed the first-time successful application *Senna macranthera* I&B., *Senna splendida* I&B. and *Senna alata* L. leaves and branches extracts as photoactive substances to treat pathogenic bacteria of *Cutibacterium acnes*, *Candida albicans*, *Staphylococcus aureus* and *Streptococcus mutans*. The study showed seven AQs and one naphthopyrone as potential photoactive substances in the extracts of liquid chromatography –diode-array detection-mass spectrometry (LC-DAD-MS) analysis [43]. The data about the microbial resistance and the treatment of viral infections including COVID-19 are in an early stage of the studies [44]. The investigation of the crude ethanolic plant extracts of *Albizia procera* (Roxb.) Benth. (Fabaceae) and *Lumnitzera*

*racemosa* Willd. (Combretaceae) showed that they are non-toxic on cancer but also on non-cancer cells [45]. However, the irradiation of the same extracts demonstrated photocytotoxic effect against breast cancer cells (MCF-7).

### 3.2 Rubiaceae (*Rubia*, *Morinda*)

Rubiaceae family contains majorly terrestrial plants including more than 13,000 species which are distributed in at least 650 genera. This family belongs to the Gentianales, which worldwide spreading makes it the fourth largest angiosperms. Rubiaceae contains a large amount of AQ derivatives, particularly in the roots [46]. The various parts of the species *Morinda elliptica* L. followed by *Heterophylleae pustulata* Hook. f. and *Morinda royoc* L. are also rich source of AQs. These very important natural products have been evaluated with various bioactivities, including antibacterial, antiviral and antitumor, and antiparasitic which make them of high interests for the research studies [47]. At the present, the increasing incidence of terminal infections caused by antibiotic-resistant bacteria tends the discovery of new drugs originated from natural substances with wide-spectrum of antimicrobial action [48].

Many Chinese herbs namely *Rheum palmatum* L., *Fallopia multiflora* Harald. and *Reynoutria japonica* Houtt. are source of AQs including aloe emodin, emodin and other biologically-active ingredients [49]. This study concluded that the development of traditional Chinese medicine based on AQs is prospective in combating the drug resistant pathogenic bacteria. The antibacterial effects of emodin, rhein and aloe emodin are more potent than those of emodin methyl ether and chrysophanol [50]. Structurally, these AQs have the same parent nucleus but they differ from their substituents on positions 3 and 6 (Fig. 2). The polar substituents  $-\text{COOH}$  and  $-\text{CH}_2\text{OH}$  as seen for rhein and aloe emodin are beneficial to antibacterial activity, while the electron-donating groups,  $-\text{CH}_3$  and  $-\text{OCH}_3$  which are in emodin and chrysophanol, and rubiadin 1-methyl ether may reduce the antibacterial activity. Obviously the structure of natural AQs for PDT has in common two or more OH groups and  $\text{OCH}_3$  substituents (Fig. 2).

Rubiaceae family of flowering plants have been reported to have a phototoxic effect under the sun-light exposure, being part of animal feedstuffs. *H. pustulata* (Rubiaceae) grows in the Andean mountain range of the northwest of Argentina, and the plants were evaluated with strong undesirable phototoxicity [51,52]. The phototoxicity of rubiadin and soranjidiol which were mostly found in the aerial parts of *H. pustulata* was observed on the animals after consuming the plant and staying on sunlight [51]. At skin level the observed effects were inflammation, erythema, itching and petechiae but also some changes of the behavior such as nervousness, aggressiveness and photophobia. The isolated AQs rubiadin 1-methyl ether, damnacanthol, soranjidiol, pustuline, damnacanthol, and heterophylline were evaluated with photosensitizing properties due to generation of superoxide anion under illumination [52]. The control group of animals kept in dark place showed neither anomalous behavior nor skin phototoxicity as well as the control group of animals exposed to light. The metabolic species were evaluated with generation of radical/ions typical for photosensitization reactions. In view of the toxicity the latest known benefit is the low systemic toxicity of AQs, which is just the opposite observation on the previous finding (excluding danthron) [51,52]. There are negligible side effects due to everyday light exposure as might be not observed for synthetic drugs.

Chemical investigation of the plant leaves and stems of *H. pustulata* revealed the numerous AQs (9,10-anthraquinone derivatives), soranjidiol, soranjidiol 1-methyl ether, rubiadin, rubiadin 1-methyl ether, damnacanthol, damnacanthol, heterophylline, pustuline, 2-hydroxy-3-methyl anthraquinone and (*S*)-5,5'-bisoranjidiol [53–55]. A study by Comini *et al.* [56] demonstrated that soranjidiol, soranjidiol 1-methyl ether, rubiadin, and rubiadin 1-methyl ether, have PDT activity on caspase-3-transfected human breast carcinoma cells (MCF-7). In this study, the percentage of cell internalization was positively correlated with the

phototoxic efficacy of the contents of AQ-compounds. The fine selection of the most effective natural AQ-sensitizer for PDT needs more experimental data.

The plant *H. pustulata* was evaluated with photoactive AQs rubiadin and soranjidiol that have different biological activity. In case of monoculture of colon cancer cells, two AQs such as soranjidiol and rubiadin showed no significant difference in the photosensitizing ability as they did in spheroid cultures [57]. It was determined that a concentration of 10  $\mu\text{g/mL}$  for rubiadin and soranjidiol retains the cellular viability up to 80% in mammalian eukaryotic Vero cells [58]. The study described that cells cultures did not deliver soranjidiol and rubiadin into the cellular nucleus, which is presenting a minimal mutagenesis risk. More so after irradiation the generation of ROS was considerably and consistently higher for the treated cells and this is one of the desirable behaviors for PSs. As well, both AQs exist as regioisomers formed during the biosynthetic shikimic acid pathway (acid *o*-succinilbenzoico; mevalonate) [59]. Photosensitizing rubiadin, soranjidiol, damnacanthol and 5,5'-bisoranjidiol isolated from *H. pustulata* Hook f. (Rubiaceae) showed antibacterial activity (bacteriostatic) on *Staphylococcus aureus* [60]. Dimmer *et al.* [61] demonstrated that irradiation enhanced the reactive oxygen production compared to the control samples without light. The mechanism of action involves an increase in the levels of superoxide anion ( $\text{O}_2^{\cdot-}$ ). Three new AQs such as lycionine (1), 7-chlorobisoranjidiol (2) and 5-chlorosoranjidiol (3) isolated from *Heterophyllaea lycioides* (Rusby) Sandwith (Rubiaceae) participate in photosensitization mechanism Type I. The study observed as an evidenced the increase of superoxide anion radical generation under the light application. The most active AQ with the greatest production of this ROS was compound 2, with an increase of 20% of this ROS compared to samples without light. The other AQs (1 and 3) produced only a slight rise of 7% ROS generation [61]. This early study also demonstrated the effect of light irradiation as an enhancing factor for the production of ROS that enhance the antibacterial activity of the applied AQ-compounds. The phenomenon is typical for PDT drugs known as prooxidant and photochemotherapeutics.

As is known the photocytotoxicity depends on the proper interactions between PS, light and oxygen, but the first object is the chemical structure of a PS, which further determines the light penetration depth and in the end the oxygen availability [62]. The oxygen atmosphere is critical for the sufficient production of ROS and other species which are needed to induce cells inactivation. The mechanism of ROS generation is typical for the photosensitive natural AQs [63]. Another study shows for the first time the reduction of biofilms of *Candida tropicalis* with two AQs [64]. More effective inactivation was observed with rubiadin (AQ1) than with rubiadin 1-methyl (AQ2). The mechanism of action after light application was examined

as mediated by an oxidative and nitrative stress with formation of endogenous ROS mainly  $O_2^{\bullet-}$  (Type I). Consequently, the prooxidant-antioxidant balance was altered specifically by AQ1. The best coactive combination with an antifungal drug Amphotericin B was also obtained with rubiadin, AQ1 (80.5%). The further study explored the generation of  $^1O_2$  (Type II) in fungal biofilms, but its contribution to the photo-initiated biofilm reduction was established as negligible [65].

### 3.3 Polygonaceae (*Rhubarb, Polygonum, Rumex*)

Polygonaceae plants have a worldwide distribution, most of which are in the temperate region of the Northern Hemisphere. There are 235 Polygonaceae species and 37 varieties in China. Plants of the Polygonaceae family have been shown with a high frequency of AQ structured compounds. The Polygonaceae are commonly known as the knotwood or smartweed family, which consist of flowering plants and comprise about 1200 species in 48 genera [66]. Among which the largest ones are *Eriogonum* Michx. (2410 spp.), *Rumex* L. (200 spp.), *Cocoloba* P. Browne (120 spp.) and *Persicaria* Mill. (100 spp.) and among others. This family is widely distributed in the North Temperature Zone although can be found worldwide [67]. The best known compounds emodin, rhein, physcion, chryso-phenol, catenarin, and pupurin have been studied to have a wide range of versatile bioactivities, including bactericidal, fungicidal, antioxidant, anti-inflammatory, anticancer, and many others attributed to substances in this family [68,69].

The well known compounds such as emodin, chryso-phenol, and aloe emodin have been isolated from different *Polygonum* spp., including *Polygonum bistorta* L. [70]. A potential antiviral activity of emodin isolated from *Rheum palmatum* L., *Rheum tanguticum* Balf., *Rheum officinale* Baill., *Polygonum cuspidatum* Siebold & Zucc. and *Polygonum multiflorum* Thunb. was reported in a recent review [13]. A traditional Chinese medicine plant *P. cuspidatum* has roots rich in three AQs namely emodin (50  $\mu\text{g}/\text{mg}$ ), physcion (10  $\mu\text{g}/\text{mg}$ ) and rhein (5.6  $\mu\text{g}/\text{mg}$ ) [71]. The results included a visible light-dependent decrease of HSV-2 plaque using the extract for inactivation of Herpes virus. The photosensitizing effect of extract was due to emodin as the main active compound with a high production of ROS while no reduction effect was observed for the control samples without light irradiation.

Many different genera of Polygonaceae family including *Rheum* spp., *Polygonum* spp. and *Rumex* spp. have been documented to contain AQ-derivatives with phototoxicity action [72]. Importantly, the antiviral activity was demonstrated to be a combination of effects of different classes of compounds with or without photoactivity. So that they can have a potential in the development of new photo-active drugs [73]. The studied AQs from the plants of the above families have shown their effects against viruses that infect the respiratory tract, including human respiratory syncytial

virus and influenza virus which infect the respiratory tract like coronavirus. AQs were evaluated with high capacity to inhibit coronavirus *in vitro* and *in silico* which could serve as a base for development of effective antiviral drugs towards coronaviruses [74].

The photodynamic activity studied for aloe emodin (AE) isolated from *Rheum palmatum* L. and *Aloe barbadensis* Miller against pathogenic bacteria and fungi showed a high efficiency of this compound [75–77]. This study suggested that the limitation of AE absorption spectrum leads to PDT effectiveness only for treatment of superficial diseases (skin cancers, infections in oral cavity, eye conditions and among others). The singlet oxygen quantum yield ( $\Phi\Delta$ ) of AE showed a value 0.54 (MeCN, 355 nm) for the studies of singlet oxygen in solution [78]. Another study of  $\Phi\Delta$  of AE determined a value 0.57 (MeOH), which is much higher than in comparison with the  $\Phi\Delta$  for phenothiazine dye toluidine blue [79]. This study also presented the cytotoxic effect of AE which was evaluated as significantly higher than that of AE applied alone without light exposure. The bioactive components in “rhubarb” are AE and emodin, both has been evaluated with favored toxicity towards carcinoma cells. Whilst it has been reported that emodin is toxic to leukemia cells only after application of blue light [80]. The cytotoxic effect gradually increased for the cells treated with different concentrations of AE and different light fluences under the identical irradiance [81]. These results indicated that AE-blue light mediated PDT offers an effective strategy for the treatment of superficial diseases without harmful effect on the normal tissues. Therefore, the authors assumed that the photoinduced damages due to PS – light interaction might be avoided. Recent study of Lin *et al.* [82] hypothesized the mechanism of AE-induced photodynamic effect on cancer cell that may be associated with apoptosis and mitochondrial pathway. The investigations of AE-photodynamic effect was carried out on human gastric cancer cells. The apoptosis and expression levels of caspases involved in the mitochondrial pathway were determined as a result of AE-PDT.

### 3.4 Asphodelaceae (*Asphodeline, Aloe, Asphodelus*)

Asphodelaceae is a family of monocotyledonous flowering plants occurring in Africa and spread from the Mediterranean to central Asia. The genus *Asphodeline* Rchb., containing 14 species, common in Mediterranean region from this family and subfamily Asphodeloideae. Three of them specifically, *A. lutea* (L.) Reichenb., *A. liburnica* (Scop.) Rchb, and *A. taurica* (Pall. ex M.Bieb.) Endl. are growing in Europe Balkan region. The main gene *Asphodeline* includes 20 taxa of which 12 are endemic in Anadolu region [83]. The *Asphodeline* spp. are traditionally used as medicinal plants in Turkish folk medicine and for human diet. These are *A. damascena* subsp. *damascena* and *A. tenuior* subsp. *tenuiflora* with applications for healing of scratches and ver-

rucae in traditional medicine [83]. The studies showed that the roots from *Asphodeline* spp. are rich source of both monomeric chrysophanol and dimeric asphodeline with probable photosensitizing effect [84]. In addition, many other AQs such as aloë-emodin, isochrysophanol, physcion, rhein, 1,5,8-trihydroxy-3-methylanthraquinone, 2-acetyl-1-hydroxy-8-methoxy-3-methylanthraquinone, asphodeline and 1,1',8,8',10-pentahydroxy-3,3'-dimethyl-10,7'-bianthracene-9,9',10'-trione were also identified [85, 86]. A chrysophanol as dimer of asphodeline and 10,7'-bichrysophanol is widely spreaded in *Asphodeline* spp., *Asphodelus* spp., *Bulbine* spp., *Bulbinella* spp., and others chrysophanol derivatives [87].

The most studied valuable genera belonging to Asphodeloideae subfamily is *Aloe vera* L. and the roots of *Aloe* spp. are containing 1-methyl-8-hydroxyanthraquinones [88]. The extracts were shown to be protective against photodamages after removing the fraction with AQ [89]. The major bioactive compound in the leaves and roots of *Aloe vera* L. is AE which has been studied with a broad spectrum of anti-cancer activity [90]. The photochemical activity of AE was evaluated with phototoxicity against the K3 cell line with an increased effect (77%) due to irradiation with blue light (405 nm, 12.8 J/cm<sup>2</sup>) but in the dark, AE was not active for concentrations below 30 µM [91]. Wamer and co-authors showed that the phototoxic effect (UVA) of AE on skin fibroblasts is mediated by oxidative damage of nucleic acids [92]. The levels of oxidized RNA were higher than DNA, which allows to speculate that AE does not accumulate in the cell nucleus. PDT with AE was shown to induce the autophagy and apoptosis of human osteosarcoma cell line MG-63 through the activation of the ROS-JNK signaling pathway [93]. Autophagy may play a protective role during the early stage following AE-PDT procedure. The further research is needed on the antitumor effect of AE-PDT to understand the modulating autophagy and prompting apoptosis.

Liu *et al.* [94] reported inhibition of human oral squamous cell carcinoma *in vitro* and *in vivo*, after application of AE-PDT. The authors concluded that AE-PDT can enhance the killing effect of KB cells (human epithelial carcinoma cells), inducing the cell apoptosis and finally leading to cell death *in vivo* and *in vitro*, hence it was evaluated as an effective therapy to cure oral cancers. It demonstrates that cell apoptosis rate in PDT is apparently higher than other single therapeutic approaches. PDT is much more effective in tumor growth inhibition and the retention of AE in tumors after treatment may promote KB cell apoptosis. This study concluded that AE mediated PDT is an innovative approach for oral cancer treatment with the supremacies of effectivity, minimal invasion, tissue integrity retention and none observed side effects on main organs. The light exposure in presence of oxygen was shown to endorse photochemical reaction including the generation of singlet oxygen but also in high extend many other ROS which can

be in advance for treatment of hypoxic pathological conditions such as malignant tumours and resistant air-free bacterial species [92–94]. The mechanism of cell death is associated with inducement of cell cycle arrest with an increment of apoptosis by upregulation of Caspase-3 and Bax proapoptotic proteins [95]. Another study on the therapeutic effect of aloë-emodin in nano-liposome (AE-L) mediated PDT showed a significant killing effect on Tca-8113 cells of human tongue squamous cell carcinoma [96]. The results indicated that for concentrations less than 7.5 µg/mL there was no significant difference between PDT group and control group ( $p > 0.05$ ). However, the increase of irradiation time led to gradual decrease the cell survival rates (76.6% for irradiation time 80 s,  $p < 0.001$ ). The results of fluorescent staining analysis showed that the reason might be due to the amount of ROS in tumor cells which increased significantly after PDT treatment. The roots of *Aloe dawei* A. Berger revealed rich content of AQs metabolite [97]. In this study two quinones 1-hydroxy-8-methoxy-3-methyl anthraquinone and 5,8-dihydroxy-3-methoxy-2-methylnaphthalene-1,4-dione were evaluated with high cytotoxicity against MCF-7 breast cancer cells. The other isolates from *A. dawei* with lesser amount of AQs were with low or medium cytotoxicity [97].

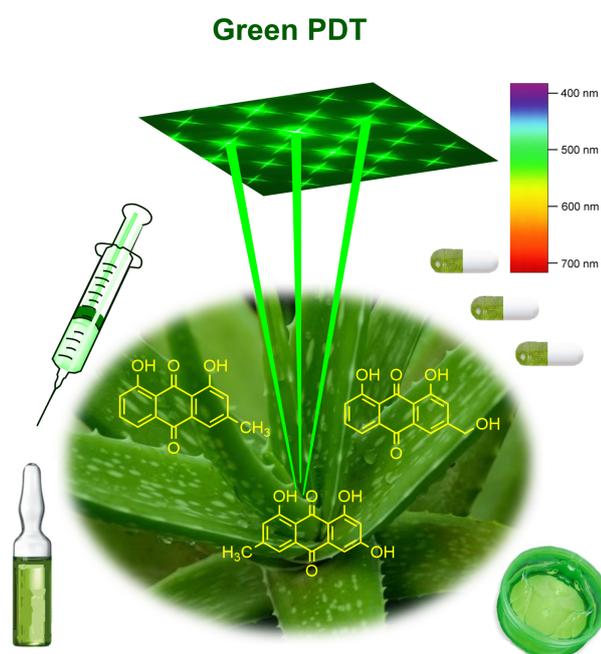
### 3.5 Rhamnaceae (*Rhamnus*, *Ventilago*)

The genus *Rhamnus* L. belongs to the Rhamnaceae family contains approximately 137 species, traditionally used in folk medicine of East Asia, North and South America, and subtropical regions of Africa. The selected species *Rhamnus purshiana* DC., *Rhamnus frangula* L., *Rhamnus catartica* L., *Ventilago maderaspatana* Gaertn. have usually been used in treatment of cancer, wound, jaundice, hepatitis, gonorrhoea, constipation, hypertension, malaria, stomach ache, snake bite and diarrhea [98]. Rhamnaceae family commonly known as a Buckthorn Family consists of ~50 genera and ~900 flowering species, mostly trees, shrubs, and some vines, and one herb. The family plants are distributed worldwide, especially in the subtropical and tropical origins [98]. The pharmacological analysis showed that the extracts and isolated several AQs found in the *Rhamnus* spp. revealed different biological activity as potential photosensitizers. The derivative prinoidin isolated from the fruits of *R. nepalensis* Laws. (Rhamnaceae) have four times more potent cytotoxic effect against human epidermoid carcinoma with IC<sub>50</sub> value of 0.045 µM than the used as a standard anticancer drug doxorubicin which is already in clinical application [99].

## 4. Discussion

The natural photosensitive compounds such as chlorophylls and chlorophyll metabolites, curcumin, thiophenes, alkaloids, as well as anthraquinones have been featured of importance for investigations as photosensitizers for PDT applications (Fig. 3). Recent work about the phototoxic

terrestrial plants including the plants containing AQs, reviews the phototoxic and photosensitizing metabolites of 250 vascular plants that have been documented as phototoxic or with photosensitive properties [100]. This review presents an in-depth investigation with the list of phytochemicals synthesized by terrestrial plants with the photosensitive compounds, and the basic molecules for the further development of new PDT-PSs. The compounds originated from different plant species are under intensive research due to their promising photocytotoxicity in the treatment of cancer and microbial infections [101]. This indicates the future for natural compounds among the class of AQs as anticancer and antiinfection therapeutics (Fig. 3). Moreover, PDT is recognized as a method with a high impact on the resistance inhibition.



**Fig. 3. Green PDT approach due to the spectra and plants' origin of the photosensitive anthraquinone compounds.**

Presently, the application of quinones as PDT agents is limited because of their intensive absorption in the whole UV spectrum, the lack of selectivity, fast clearance and strong hydrophobicity. However, there are few derivatives that can be used as potential PDT agents after properly performed structural modification, so that the absorption relocates to the longer wavelength ( $>500$  nm), improves selectivity and also reduces hydrophobicity [102]. The positive aspect of AQs' mechanism of action is their ability to produce toxic species without loss of cytotoxic abilities in pathologic sites with a low oxygen level, such as hypoxic tumors or oxygen-free pathogenic bacteria [103]. Moreover, the production of the reactive species might happen without light exposure of AQs. The appropriate photosensitization mechanisms responsible for PDT outcome require a

sufficient oxygen surrounding atmosphere. The early studies suggested that the natural AQ compounds such as aloe emodin and hypericin can be considered as promising PS candidates for PDT [104]. The recent advances in anticancer activity of emodin, aloe emodin, hypericin, chryso-phenol, rhein, and physcion showed these AQs as effective individually but also in combination with other chemotherapeutic agents on *in vitro* and *in vivo* basal carcinoma (BC) models [105].

Currently, several chemotherapeutic anticancer agents containing AQ-nucleus are the Food and Drug Administration (FDA) approved drugs, namely daunorubicin, idarubicin, doxorubicin (DOX), epirubicin, valrubicin, pixantrone and mitoxantrone all for clinical use in cancer therapy [106]. The appearance of drug resistance and some possible side effects as for example, the heart muscle injury after DOX treatment, lower the use of doxorubicin-based cancer treatments. Mechanisms of antitumor action of these anthracycline drugs are associated with hyperproduction of ROS and inhibition of glycolysis [107]. However, there is an emerging limitation related to the poor efficacy of chemotherapeutic agents, due to the resistance of tumor cells to these drugs, as well as due to toxicity-related side effects [108]. The score of new drug discovery is known as very low with the estimation that only ten of ten thousand novel tested compounds have been passed the regulations to enter clinical stage, and then only one may be approved through regulatory valuation and licensed for the medical application. The biologically active substances require a long term and complicated procedure including many stages of research, pre- and clinical trials on humans to develop medicine for the pharmacy shelf [109].

Gollnick *et al.* [110] (1992) showed in an initial study on AQs that the compounds bearing hydroxyl groups in their structure which are distributed on the benzene rings of both sides may have promising potential for photodynamic action. The further study on the photosensitizing properties of some natural AQs isolated from leaves and stems of *Heterophyllaea pustulata* Hook. f. (Rubiaceae) showed the both mechanisms of photosensitization for the studied rubiadin, soranjidiol, soranjidiol-1-methyl ether, damnacanthol and damnacanthol [111]. The photosensitized generation of superoxide anion radical (Type I mechanism) was presented only for rubiadin-1-methyl ether as was determined in leukocyte suspensions. The singlet oxygen generation (Type II mechanism) was studied in solution for damnacanthol (0.70) followed by soranjidiol (0.47) which were evaluated with relatively very high yields.

Although there are drugs with AQ structure that have already been approved for clinical practice, a large number of AQ analogues have been synthesized and the synthesis continued [112]. The structure of AQ is suitable for metal/non-metal coordination, but there is a limited number of studies on synthetic AQs prepared as a complex [113]. The coordination of metals was studied with the synthetic

ligands through the carbonyl group at AQ-ring and nitrogen atom of amide group. The antibacterial inactivation of both Gram-positive and Gram-negative pathogenic bacterial species showed the high activity for complexes types [Cd(L1)2Cl2] and [Cd(L2)2Cl2]. The clinical AQs have been found to have positive cardiotoxicity, but after the formation of complexes with metals such as Cu, Co, Fe and Mn was shown to limit the ROS production and reduce their toxic and side effects [114]. This can be a starting point for design of novel AQ complexes with low side effect and the superior mechanism Type II photosensitization for PDT.

A very recent study summarized the phototoxic effect of AQs and the plants of their origin and the compounds that are with promising efficiency on different experimental cancer cells [115]. The research and review papers present the AQs such as emodin, aloe-emodin, parietin, rubiadin, hypericin, and soranjidiol as photosensitive compounds for cancer PDT [116,117].

Generally, depending on drug and light administration conditions, the PDT effect induces the cell death through several mechanisms, which can be the initiation of apoptosis, autophagy-associated cell death or necrosis [118,119]. Another possible mechanism could be an immunogenic cell death (ICD) [120]. A recent study on aloe emodin (AE) from *Rheum palmatum*'s rhizome reported a mechanism of the PDT of apoptosis in target cells [121]. The mechanisms of cell-death have been mostly investigated for the natural hypericin [122]. Hypericin shows absorption with maximum close to therapeutic region (630 nm–850 nm), which by the irradiation leads to generation of ROS as well as singlet oxygen. In cells, HYP primarily accumulates in the hydrophobic core of the membrane by forming non-covalent persistent interactions and the lipid double bonds, plus in the endoplasmic reticulum (ER), mitochondria, lysosomes and Golgi apparatus [123]. Recently, Kessel reported the capacity of HYP to target the ER for photodamage, which provokes a mainly unexplored manner of photokilling that implicates extensive cytoplasmic vacuole formation but does not represent autophagy [124]. This phenomenon called ‘paraptosis’ seems to be a response to the appearance of misfolded ER proteins as important to eradicate the malignant cell population with a reduced apoptotic pathway [125,126].

A review paper about “anthraquinones as a promising scaffold for the discovery and development of therapeutic agents in cancer therapy” concluded that *in silico* approaches (molecular modeling and docking studies) have been recognized as an extensively useful support for progress of novel anticancer compounds based on AQ-moieties [127,128]. The knowledge on the binding interactions of AQs with amino acids located in active site pocket of targeting enzymes such as CK2 $\alpha$ , topoisomerase II  $\beta$ , human Ser/Thr kinase pim1 and reacted quinone reductase might lead to further promising modifications [128]. The AQ core molecule gives the idea to develop an advantaged

scaffold in medicinal chemistry in targeting various biological proteins. The genetic reports including topoisomerases, telomerase, protein kinases and matrix metalloproteinases (MMPs) have been widely highlighted. Since the natural AQs and their synthetic derivatives show an inclusive variety of biological activities. Some of them are with high social importance suggesting their probable medical application and surely might serve as a source of effective medications for the progress in development of anticancer drugs [129].

The conformational analysis with three-dimensional quantitative structure-activity relationships (3D-QSAR) of compounds and thus the design of new synthetic photosensitizers could provide a methodological reference for the design and synthesis studies of novel photosensitizers [130,131]. The AQ moiety provides a potential substrate for DNA insertion and bio-reduction activation, and the alkylation group confers cytotoxic activity [130]. DNA intercalators that typically contain coplanar aromatic backbones such as doxorubicin, daunorubicin, and mitoxantrone (MTZ) can be reversibly inserted into base pairs of DNA. The scientists reported that AQ moieties may serve as a base for preparation of synthetic hydroxyanthraquinone derivatives as the recently reported with *N*-mustards, thiophene and benzene ring as substituents. The molecular docking studies of compounds demonstrated that the new compound (A1) could interact with the catalytic active site in topoisomerase II. The recent paper studied five pH-responsive anthraquinone photosensitizers (A1 to A5) synthesized through addition, elimination, and substitution mechanisms using chrysophanol as the raw material [131]. The compounds were shown to have good PDT antibacterial activity and selectivity in weak acid environments. The cytotoxicity (3-[4,5-dimethylthiazol-2-yl]-2,5 diphenyl tetrazolium bromide, MTT test) on the cancer cell line HeLa demonstrated the photo- and dark toxicity effects of compounds (for A3 at 0.625  $\mu$ M) similar to the reference photosensitizer Chlorin e6 (Ce6). Nevertheless these findings, it is still very questionable to find a very powerful PS among the plants-originated AQ-compounds because of the spectral properties of short-wavelength absorption (<600 nm). One obvious explanation of the low toxicity and gentle photodynamic action after irradiation can be explained with the evolution stage under the sun light and oxygen atmosphere, and both are involved in photosensitization but also they are critical for the plants' growth and their being in material sense.

## 5. Perspectives of Anthraquinones for PDT

Green approach to PDT includes the PS isolated from plants or other natural source and the specific visible light spectra (400–600 nm), and sufficient oxygen surrounding [132]. Presently, there are no clinically approved plants isolated photosensitizers (PSs) for application as drugs in medical PDT. It is only hypericin which pass throughout

many years of research and development and pre-clinical studies so that it has been featured with good perspectives for clinical applications [133]. The scientific society expect that it is the near future when the natural PSs and their synthetic analogues to be invented in cancer PDT. The natural AQs can serve as a core molecule for design of novel more powerful anticancer agents with photosensitive properties that are tailored for PDT.

Practically the natural compounds exhibited the poor solubility, high body clearance rate and restricted tumor targeting capability together the expected lower efficiency as PDT drugs. In order to solve these problems, natural PSs and their derivatives might be involved in molecular assembly technique to enhance uptake, selectivity and PDT efficacy in addition to minimal toxicity in the presence of visible light as the main advantage of these PSs. Also, since natural compounds are usually universal, and some of them are already in usage as herbs and therapeutic drugs, they may be more readily-accessible and easy for therapeutic invention.

In future, researchers may discover these photoactive plants in nature and compare the effects with clinical drugs. Furthermore, owning on the well-accepted structures of PSs for PDT, the natural AQs' might serve as a template for development of variety of novel target-specific PSs containing definite functional groups as substituents for the target specific PDT. The novel compounds may have the tunability of the photochemical and the photobiological properties, high biocompatibility, and selective cellular uptake in the tumors or other target pathologic tissue and cells, and not the last the better solubility in natural fluids, and absorbance in the optical PDT window (630–850 nm). In addition, the far-red fluorescence emission will be advantageous for the non-invasive optical diagnosis.

## 6. Conclusions

As researchers, we must consider the enquiry whether or not is realistic the anthraquinone—associated PDT to get permission for clinical practice? The crucial aspect about plants' PS is that they have not characteristics of typical photosensitizers due to the natural exposure to sunlight which could be harmful and destructive during the plants growing. However, there are good opportunities for natural anthraquinone hypericin, as it is already on clinical level in cancer PDT. Since the progress in PDT involves the applications of the method to the actual diseases due to resistance to present drugs and for the other diseases without alternative treatments or with insufficient treatment consequence. The decisive stage for further development of natural plant-occurring AQ for PDT applications is to find the unique plant or plants that are having high quantity appropriate compounds and substances with valuable photosensitizing properties. This fact will allow to develop a procedure to obtain the photoactive compound(s) in a large scale. Moreover, the plant cultures hold great potential for the manu-

facture of phytochemicals to obtain natural compounds on a large scale, in an economically viable and eco-friendly manner.

## Abbreviations

AQ, anthraquinones; AE, aloe emodin; HYP, hypericin; ROS, reactive-oxygen species; LED, light-emitting diode; PS, photosensitizers; PDT, photodynamic therapy.

## Author Contributions

VM designed, made the searches, wrote original draft, made the corrections, supervised, and answered the reviewers. DB made the searches, wrote Supplementary, prepared original Figures and made critical corrections. IL made searches, wrote paragraphs, and reviewed. TG made searches, wrote draft, and made the critical corrections. All authors read and approved the final manuscript. All authors have participated sufficiently in the work and agreed to be accountable for all aspects of the work.

## Ethics Approval and Consent to Participate

Not applicable.

## Acknowledgment

Thanks to Bulgarian Academy of Sciences.

## Funding

This research received no external funding.

## Conflict of Interest

Given her role as Guest Editor, Vanya Mantareva had no involvement in the peer-review of this article and has no access to information regarding its peer review. Full responsibility for the editorial process for this article was delegated to Jen-Tsung Chen. The authors declare no conflict of interest.

## Supplementary Material

Supplementary material associated with this article can be found, in the online version, at <https://doi.org/10.31083/j.fbl2905168>.

## References

- [1] Kubrak TP, Kołodziej P, Sawicki J, Mazur A, Koziarowska K, Aebischer D. Some Natural Photosensitizers and Their Medicinal Properties for Use in Photodynamic Therapy. *Molecules* (Basel, Switzerland). 2022; 27: 1192.
- [2] Cimino F, Saija A, Speciale A. Chapter 13 - Applications of plant-derived products in photodynamic therapy. In Saso L, Giuffrè A, Valacchi G, Maccarrone M (eds.) *Modulation of Oxidative Stress*. (pp. 175–197). Academic Press: London. 2023.
- [3] Gonçalves LCP. Photophysical properties and therapeutic use of natural photosensitizers. *Journal of Photochemistry and Photobiology*. 2021; 7: 100052.
- [4] Malik EM, Müller CE. Anthraquinones As Pharmacological

- Tools and Drugs. *Medicinal Research Reviews*. 2016; 36: 705–748.
- [5] Aziz B, Aziz I, Khurshid A, Raoufi E, Esfahani FN, Jalilian Z, *et al.* An Overview of Potential Natural Photosensitizers in Cancer Photodynamic Therapy. *Biomedicines*. 2023; 11: 224.
  - [6] Penetra M, Arnaut LG, Gomes-da-Silva LC. Trial watch: an update of clinical advances in photodynamic therapy and its immunoadjuvant properties for cancer treatment. *Oncoimmunology*. 2023; 12: 2226535.
  - [7] Choudhary N, Collignon TE, Tewari D, Bishayee A. Hypericin and its anticancer effects: From mechanism of action to potential therapeutic application. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2022; 105: 154356.
  - [8] Li X, Liu Y, Wu L, Zhao J. Molecular Nanoarchitectonics of Natural Photosensitizers and Their Derivatives Nanostructures for Improved Photodynamic Therapy. *ChemMedChem*. 2024; 19: e202300599.
  - [9] Polat E, Kang K. Natural Photosensitizers in Antimicrobial Photodynamic Therapy. *Biomedicines*. 2021; 9: 584.
  - [10] Dave H, Ledwani L. A review on anthraquinones isolated from Cassia species and their applications. *Indian Journal of Natural Products and Resources*. 2012; 3: 291–319.
  - [11] Christie R, Abel A. Anthraquinonoid pigments. *Physical Sciences Reviews*. 2021; 6: 299–313.
  - [12] Duval J, Pecher V, Poulou M, Lesellier E. Research advances for the extraction, analysis and uses of anthraquinones: A review. *Industrial Crops and Products*. 2016; 94: 812–833.
  - [13] Ntemafack A, Singh RV, Ali S, Kuate JR, Hassan QP. Antiviral potential of anthraquinones from Polygonaceae, Rubiaceae and Asphodelaceae: Potent candidates in the treatment of SARS-COVID-19, A comprehensive review. *South African Journal of Botany: Official Journal of the South African Association of Botanists = Suid-Afrikaanse Tydskrif Vir Plantkunde: Amptelike Tydskrif Van Die Suid-Afrikaanse Genootskap Van Plantkundiges*. 2022; 151: 146–155.
  - [14] Dimmer J, Cabral FV, Sabino CP, Silva CR, Núñez-Montoya SC, Cabrera JL, *et al.* Natural anthraquinones as novel photosensitizers for antiparasitic photodynamic inactivation. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2019; 61: 152894.
  - [15] Liang R, Wong KH, Yang Y, Duan Y, Chen M. ROS-responsive dexamethasone micelles normalize the tumor microenvironment enhancing hypericin in cancer photodynamic therapy. *Biomaterials Science*. 2022; 10: 1018–1025.
  - [16] Wu JJ, Zhang J, Xia CY, Ding K, Li XX, Pan XG, *et al.* Hypericin: A natural anthraquinone as promising therapeutic agent. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2023; 111: 154654.
  - [17] Zhang JN, Zhang F, Tang QJ, Xu CS, Meng XH. Effect of photodynamic inactivation of *Escherichia coli* by hypericin. *World Journal of Microbiology & Biotechnology*. 2018; 34: 100.
  - [18] Rumie Vittar NB, Comini L, Fernandez IM, Agostini E, Nuñez-Montoya S, Cabrera JL, *et al.* Photochemotherapy using natural anthraquinones: Rubiadin and Soranjidiol sensitize human cancer cell to die by apoptosis. *Photodiagnosis and Photodynamic Therapy*. 2014; 11: 182–192.
  - [19] George BP, Abrahamse H. Light-Activated Phytochemicals in Photodynamic Therapy for Cancer: A Mini Review. *Photobiomodulation, Photomedicine, and Laser Surgery*. 2022; 40: 734–741.
  - [20] Aires-Fernandes M, Botelho Costa R, Rochetti do Amaral S, Mussagy CU, Santos-Ebinuma VC, Primo FL. Development of Biotechnological Photosensitizers for Photodynamic Therapy: Cancer Research and Treatment-From Benchtop to Clinical Practice. *Molecules (Basel, Switzerland)*. 2022; 27: 6848.
  - [21] Danta CC, Sahu AN. Chapter 16 - Naturally occurring anticancer drugs. In Pratap CA, Kurosu M (eds.) *Medicinal Chemistry of Chemotherapeutic Agents* (pp. 539–588). Academic Press: Amsterdam. 2023.
  - [22] Swedan HK, Kassab AE, Gedawy EM, Elmeligie SE. Topoisomerase II inhibitors design: Early studies and new perspectives. *Bioorganic Chemistry*. 2023; 136: 106548.
  - [23] van der Zanden SY, Qiao X, Neeffjes J. New insights into the activities and toxicities of the old anticancer drug doxorubicin. *The FEBS Journal*. 2021; 288: 6095–6111.
  - [24] Tikhomirov AS, Tsvetkov VB, Volodina YL, Litvinova VA, Andreeva DV, Dezhenkova LG, *et al.* Heterocyclic ring expansion yields anthraquinone derivatives potent against multidrug resistant tumor cells. *Bioorganic Chemistry*. 2022; 127: 105925.
  - [25] Sokkar P, Babu A, Kolandaswamy A, Daison FA, Ramachandran M. Effect of Substituents on the Photodynamic Action of Anthraquinones: EPR, Computational and In Vitro Studies. *Photochemistry and Photobiology*. 2022; 98: 1426–1433.
  - [26] Diaz-Muñoz G, Miranda IL, Sartori SK, de Rezende DC, Diaz MAN. Chapter 11 - Anthraquinones: An Overview. In Rahman A (ed.) *Studies in Natural Products Chemistry* (pp. 313–338). Elsevier: Amsterdam. 2018.
  - [27] Yadav AN, Kour D, Rana KL, Yadav N, Singh B, Chauhan VS, *et al.* Chapter 20 - Metabolic Engineering to Synthetic Biology of Secondary Metabolites Production. In Gupta VK, Pandey A (eds.) *New and Future Developments in Microbial Biotechnology and Bioengineering* (pp. 279–320). Elsevier: Cambridge. 2019.
  - [28] Nuñez Montoya SC, Agnese AM, Pérez C, Tiraboschi IN, Cabrera JL. Pharmacological and toxicological activity of *Heterophyllaea pustulata* anthraquinone extracts. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2003; 10: 569–574.
  - [29] Nuñez Montoya SC, Agnese AM, Cabrera JL. Anthraquinone derivatives from *Heterophyllaea pustulata*. *Journal of Natural Products*. 2006; 69: 801–803.
  - [30] Caro Y, Anamale L, Fouillaud M, Laurent P, Petit T, Dufossé L. Natural hydroxyanthraquinoid pigments as potent food grade colorants: An overview. *Natural Products and Bioprospecting*. 2012; 2: 174–193.
  - [31] Murthy HN, Joseph KS, Paek KY, Park SY. Anthraquinone Production from Cell and Organ Cultures of *Rubia* Species: An Overview. *Metabolites*. 2022; 13: 39.
  - [32] Ziani BEC, Mohamed A, Ziani C, Saher L. Polyketides. In Carcho M, Heleno SA, Barros L (eds.) *Natural Secondary Metabolites*. Springer, Cham: Cham. 2023.
  - [33] Deshpande HA and Bhalsing SR. Recent advances in the phytochemistry of some medically important Cassia species: A REVIEW. *International Journal of Pharma Medicine and Biological Sciences*. 2013; 2: 60–78.
  - [34] Cai Z, Zhao X, Zhou C, Fang T, Liu G, Luo J. Genome-Wide Mining of the Tandem Duplicated Type III Polyketide Synthases and Their Expression, Structure Analysis of *Senna tora*. *International Journal of Molecular Sciences*. 2023; 24: 4837.
  - [35] Wilson L. Spices and Flavoring Crops: Leaf and Floral Structures. In Benjamin C, Paul MF, Fidel T (eds.) *Encyclopedia of Food and Health* (pp. 84–92). Academic Press: Waltham. 2016.
  - [36] Awomukwu AD, Nyananyo BL, Ikepeama AI, Adieze CU. Comparative Chemical Constituents of some Cassia Species and their Pharmacognostic Importance in South Eastern Nigeria. *Science Journal of Chemistry*. 2015; 3: 40–49.
  - [37] Singh S, Singh SK, Yadav A. A Review on Cassia species: Pharmacological, Traditional and Medicinal Aspects in Various Countries. *American Journal of Phytomedicine and Clinical Therapeutics*. 2013; 3: 291–312.
  - [38] Maroyi A. Medicinal Uses of the Fabaceae Family in Zimbabwe: A Review. *Plants (Basel, Switzerland)*. 2023; 12: 1255.

- [39] Sharma A, Flores-Vallejo RDC, Cardoso-Taketa A, Villarreal ML. Antibacterial activities of medicinal plants used in Mexican traditional medicine. *Journal of Ethnopharmacology*. 2017; 208: 264–329.
- [40] Bee BS, Jat RK, Ahmad S. Fabrication of microspheres and characterization of antimicrobial and anti-inflammatory activity isolated fraction from total alcoholic extract of *Cassia Fistula* (Linn.) in carrageenan-Type-IV induced inflammatory rats. *International Journal of Experimental Research and Review*. 2023; 32: 246–259.
- [41] Zhang S, Li X, Lu X, Long C, Zhao Y, Yu Z. Structure and Bioactivity of Purified Polysaccharides of *Cassia occidentalis* Linn. from Different Geographical Origins. *Journal of Polymers and the Environment*. 2023; 31: 5137–5151.
- [42] Konar A, Gosh P, Chatterjee S. *Cassia fistula* is a Miraculous Medicinal Plant: A Brief Review. *Sarcouncil Journal of Plant and Agronomy*. 2023; 1: 25–31.
- [43] Oliveira ABD, Ferrisse TM, Annunzio SRD, Franca MGA, Silva MGDV, Cavalheiro AJ, *et al.* In Vitro Evaluation of Photodynamic Activity of Plant Extracts from *Senna* Species against Microorganisms of Medical and Dental Interest. *Pharmaceutics*. 2023; 15: 181.
- [44] Pattnaik S, Mishra M, Naik PK. Alternative Strategies for Combating Antibiotic Resistance in Microorganisms. *Antimicrobial Photodynamic Therapy* (pp. 45). CRC Press: Boca Raton. 2023.
- [45] Villacorta RB, Roque KFJ, Tapang GA, Jacinto SD. Plant extracts as natural photosensitizers in photodynamic therapy: in vitro activity against human mammary adenocarcinoma MCF-7 cells. *Asian Pacific Journal of Tropical Biomedicine*. 2017; 7: 358–366.
- [46] Martins D, Nunez CV. Secondary metabolites from Rubiaceae species. *Molecules* (Basel, Switzerland). 2015; 20: 13422–13495.
- [47] Su GY, Chen ML, Wang KW. Natural New Bioactive Anthraquinones from Rubiaceae. *Mini-Reviews in Organic Chemistry*. 2020; 17: 872–883.
- [48] Jadimurthy R, Jagadish S, Nayak SC, Kumar S, Mohan CD, Rangappa KS. Phytochemicals as Invaluable Sources of Potent Antimicrobial Agents to Combat Antibiotic Resistance. *Life* (Basel, Switzerland). 2023; 13: 948.
- [49] Qun T, Zhou T, Hao J, Wang C, Zhang K, Xu J, *et al.* Antibacterial activities of anthraquinones: structure-activity relationships and action mechanisms. *RSC Medicinal Chemistry*. 2023; 14: 1446–1471.
- [50] Friedman M, Xu A, Lee R, Nguyen DN, Phan TA, Hamada SM, *et al.* The Inhibitory Activity of Anthraquinones against Pathogenic Protozoa, Bacteria, and Fungi and the Relationship to Structure. *Molecules* (Basel, Switzerland). 2020; 25: 3101.
- [51] Núñez Montoya SC, Comini LR, Rumie Vittar B, Fernández IM, Rivarola VA, Cabrera JL. Phototoxic effects of *Heterophyllaea pustulata* (Rubiaceae). *Toxicon: Official Journal of the International Society on Toxinology*. 2008; 51: 1409–1415.
- [52] Micheloud JF, Aguirre LS, Marioni J, Mugas ML, Cabrera JL, Martinez OG, *et al.* Experimental poisoning by *Heterophyllaea pustulata* Hook. f. (Rubiaceae) in goats. Clinical, biochemical and toxicological aspects. *Toxicon: Official Journal of the International Society on Toxinology*. 2019; 165: 56–61.
- [53] Watroly MN, Sekar M, Fuloria S, Gan SH, Jeyabalan S, Wu YS, *et al.* Chemistry, Biosynthesis, Physicochemical and Biological Properties of Rubiadin: A Promising Natural Anthraquinone for New Drug Discovery and Development. *Drug Design, Development and Therapy*. 2021; 15: 4527–4549.
- [54] Luo H, Qin W, Zhang H, Ren FC, Fang WT, Kong QH, *et al.* Anthraquinones from the Aerial Parts of *Rubia cordifolia* with Their NO Inhibitory and Antibacterial Activities. *Molecules* (Basel, Switzerland). 2022; 27: 1730.
- [55] Mansoori B, Mohammadi A, Amin Doustvandi M, Mohammadnejad F, Kamari F, Gjerstorff MF, *et al.* Photodynamic therapy for cancer: Role of natural products. *Photodiagnosis and Photodynamic Therapy*. 2019; 26: 395–404.
- [56] Comini LR, Fernandez IM, Rumie Vittar NB, Núñez Montoya SC, Cabrera JL, Rivarola VA. Photodynamic activity of anthraquinones isolated from *Heterophyllaea pustulata* Hook f. (Rubiaceae) on MCF-7c3 breast cancer cells. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2011; 18: 1093–1095.
- [57] Cogno IS, Gilardi P, Comini L, Núñez-Montoya SC, Cabrera JL, Rivarola VA. Natural photosensitizers in photodynamic therapy: In vitro activity against monolayers and spheroids of human colorectal adenocarcinoma SW480 cells. *Photodiagnosis and Photodynamic Therapy*. 2020; 31: 101852.
- [58] Montoya SC, Comini LR, Cabrera JL. Antimicrobial activity of natural photosensitizing anthraquinones. *Science against Microbial Pathogens: communicating current research and technological advances A*. 2011; 3–13.
- [59] Talapatra SK, Talapatra B. Shikimic Acid Pathway. *Chemistry of Plant Natural Products* (pp. 625–678). Springer: Berlin, Heidelberg. 2015.
- [60] Comini LR, Montoya SCN, Páez PL, Argüello GA, Albesa I, Cabrera JL. Antibacterial activity of anthraquinone derivatives from *Heterophyllaea pustulata* (Rubiaceae). *Journal of Photochemistry and Photobiology. B, Biology*. 2011; 102: 108–114.
- [61] Dimmer JA, Núñez Montoya SC, Mendoza CS, Cabrera JL. Photosensitizing anthraquinones from *Heterophyllaea lycioides* (Rubiaceae). *Phytochemistry*. 2017; 137: 94–100.
- [62] Tian W, Wang C, Li D, Hou H. Novel anthraquinone compounds as anticancer agents and their potential mechanism. *Future Medicinal Chemistry*. 2020; 12: 627–644.
- [63] Malmir M, Serrano R, Duarte Silva OM. Anthraquinones as potential antimicrobial agents-A review. In Mendez-Vilas A (ed.) *Antimicrobial research: Novel bio-knowledge and educational programs* (pp. 55–61). Formatex: Badajoz. 2017.
- [64] Marioni J, da Silva MA, Cabrera JL, Montoya SCN, Paraje MG. The anthraquinones rubiadin and its 1-methyl ether isolated from *Heterophyllaea pustulata* reduces *Candida tropicalis* biofilms formation. *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2016; 23: 1321–1328.
- [65] Marioni J, Bresoli-Obach R, Agut M, Comini LR, Cabrera JL, Paraje MG, *et al.* On the mechanism of *Candida tropicalis* biofilm reduction by the combined action of naturally-occurring anthraquinones and blue light. *PloS One*. 2017; 12: e0181517.
- [66] Xu Z, Deng M. Polygonaceae. *Identification and Control of Common Weeds*. 2017; 2.
- [67] Li JJ, Li YX, Li N, Zhu HT, Wang D, Zhang YJ. The genus *Rumex* (Polygonaceae): an ethnobotanical, phytochemical and pharmacological review. *Natural Products and Bioprospecting*. 2022; 12: 21.
- [68] Hochma E, Yarmolinsky L, Khalfin B, Nisnevitch M, Ben-Shabat S, Nakonechny F. Antimicrobial Effect of Phytochemicals from Edible Plants. *Processes*. 2021; 9: 2089.
- [69] Prateeksha, Yusuf MA, Singh BN, Sudheer S, Kharwar RN, Siddiqui S, *et al.* Chrysophanol: A Natural Anthraquinone with Multifaceted Biotherapeutic Potential. *Biomolecules*. 2019; 9: 68.
- [70] Prakash Mishra A, Sharifi-Rad M, Shariati MA, Mabkhot YN, Al-Showiman SS, Rauf A, *et al.* Bioactive compounds and health benefits of edible *Rumex* species-A review. *Cellular and Molecular Biology (Noisy-le-Grand, France)*. 2018; 64: 27–34.
- [71] Ntemafack A, Ayoub M, Hassan QP, Gandhi SG. A systematic review of pharmacological potential of phytochemicals from *Rumex abyssinicus* Jacq. *South African Journal of Botany*. 2023; 154: 11–25.

- [72] Monjo ALA, Pringle ES, Thornbury M, Duguay BA, Monro SMA, Hetu M, *et al.* Photodynamic Inactivation of Herpes Simplex Viruses. *Viruses*. 2018; 10: 532.
- [73] Shen BB, Yang YP, Yasamin S, Liang NA, Su W, Chen SH, *et al.* Analysis of the Phytochemistry and Bioactivity of the Genus *Polygonum* of Polygonaceae. *Digital Chinese Medicine*. 2018; 1: 19–36.
- [74] Scotti F, Mou L, Huang C, Booker A, Weckerle C, Maake C, *et al.* Treating Chronic Wounds Using Photoactive Metabolites: Data Mining the Chinese Pharmacopoeia for Potential Lead Species. *Planta Medica*. 2021; 87: 1206–1218.
- [75] Jamal QMS. Antiviral Potential of Plants against COVID-19 during Outbreaks-An Update. *International Journal of Molecular Sciences*. 2022; 23: 13564.
- [76] Otieno W, Liu C, Ji Y. Aloe-Emodin-Mediated Photodynamic Therapy Attenuates Sepsis-Associated Toxins in Selected Gram-Positive Bacteria In Vitro. *Journal of Microbiology and Biotechnology*. 2021; 31: 1200–1209.
- [77] Li J, Qin M, Liu C, Ma W, Zeng X, Ji Y. Antimicrobial photodynamic therapy against multidrug-resistant *Acinetobacter baumannii* clinical isolates mediated by aloe-emodin: An in vitro study. *Photodiagnosis and Photodynamic Therapy*. 2020; 29: 101632.
- [78] Wang Y, Li J, Geng S, Wang X, Cui Z, Ma W, *et al.* Aloe-emodin-mediated antimicrobial photodynamic therapy against multidrug-resistant *Acinetobacter baumannii*: An in vivo study. *Photodiagnosis and Photodynamic Therapy*. 2021; 34: 102311.
- [79] Ma W, Liu C, Li J, Hao M, Ji Y, Zeng X. The effects of aloe emodin-mediated antimicrobial photodynamic therapy on drug-sensitive and resistant *Candida albicans*. *Photochemical & Photobiological Sciences: Official Journal of the European Photochemistry Association and the European Society for Photobiology*. 2020; 19: 485–494.
- [80] Vath P, Wamer WG, Falvey DE. Photochemistry and phototoxicity of aloe emodin. *Photochemistry and Photobiology*. 2002; 75: 346–352.
- [81] Zang L, Zhao H, Ji X, Cao W, Zhang Z, Meng P. Photophysical properties, singlet oxygen generation efficiency and cytotoxic effects of aloe emodin as a blue light photosensitizer for photodynamic therapy in dermatological treatment. *Photochemical & Photobiological Sciences: Official Journal of the European Photochemistry Association and the European Society for Photobiology*. 2017; 16: 1088–1094.
- [82] Lin HD, Li KT, Duan QQ, Chen Q, Tian S, Chu ESM, *et al.* The effect of aloe-emodin-induced photodynamic activity on the apoptosis of human gastric cancer cells: A pilot study. *Oncology Letters*. 2017; 13: 3431–3436.
- [83] Zengin G, Locatelli M, Ceylan R, Aktumsek A. Anthraquinone profile, antioxidant and enzyme inhibitory effect of root extracts of eight Asphodeline taxa from Turkey: can Asphodeline roots be considered as a new source of natural compounds? *Journal of Enzyme Inhibition and Medicinal Chemistry*. 2016; 31: 754–759.
- [84] Zengin G, Locatelli M, Ferrante C, Menghini L, Orlando G, Brunetti L, *et al.* New pharmacological targets of three Asphodeline species using in vitro and ex vivo models of inflammation and oxidative stress. *International Journal of Environmental Health Research*. 2019; 29: 520–530.
- [85] Lazarova I, Zengin G, Gevrenova R, Nedialkov P, Aneva I, Aumeeruddy MZ, *et al.* A comparative study of UH-PLC/Orbitrap MS metabolomics profiles and biological properties of *Asphodeline taurica* from Bulgaria and Turkey. *Journal of Pharmaceutical and Biomedical Analysis*. 2019; 168: 174–180.
- [86] Lazarova I, Simeonova R, Vitcheva V, Kondeva-Burdina M, Gevrenova R, Zheleva-Dimitrova D, *et al.* Hepatoprotective and antioxidant potential of *Asphodeline lutea* (L.) Rchb. roots extract in experimental models in vitro/in vivo. *Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie*. 2016; 83: 70–78.
- [87] Wyk BEV, Yenesew A, Dagne E. Chemotaxonomic significance of anthraquinones in the roots of Asphodeloideae (Asphodelaceae). *Biochemical Systematics and Ecology*. 1995; 23: 277–281.
- [88] Raksha B, Pooja S, Babu S. Bioactive Compounds and Medicinal Properties of Aloe Vera L.: An Update. *Journal of Plant Sciences*. 2014; 2: 102–107.
- [89] Rodrigues D, Viotto AC, Checchia R, Gomide A, Severino D, Itri R, *et al.* Mechanism of Aloe Vera extract protection against UVA: shelter of lysosomal membrane avoids photodamage. *Photochemical & Photobiological Sciences: Official Journal of the European Photochemistry Association and the European Society for Photobiology*. 2016; 15: 334–350.
- [90] Chen R, Zhang J, Hu Y, Wang S, Chen M, Wang Y. Potential antineoplastic effects of Aloe-emodin: a comprehensive review. *The American Journal of Chinese Medicine*. 2014; 42: 275–288.
- [91] Siewert B, Stuppner H. The photoactivity of natural products - An overlooked potential of phytomedicines? *Phytomedicine: International Journal of Phytotherapy and Phytopharmacology*. 2019; 60: 152985.
- [92] Wamer WG, Vath P, Falvey DE. In vitro studies on the photobiological properties of aloe emodin and aloin A. *Free Radical Biology & Medicine*. 2003; 34: 233–242.
- [93] Tu P, Huang Q, Ou Y, Du X, Li K, Tao Y, *et al.* Aloe-emodin-mediated photodynamic therapy induces autophagy and apoptosis in human osteosarcoma cell line MG-63 through the ROS/JNK signaling pathway. *Oncology Reports*. 2016; 35: 3209–3215.
- [94] Liu YQ, Meng PS, Zhang HC, Liu X, Wang MX, Cao WW, *et al.* Inhibitory effect of aloe emodin mediated photodynamic therapy on human oral mucosa carcinoma in vitro and in vivo. *Biomedicine & Pharmacotherapy = Biomedecine & Pharmacotherapie*. 2018; 97: 697–707.
- [95] Alarcón J, Cespedes CL. Chemical constituents and biological activities of South American Rhamnaceae. *Phytochemistry Reviews*. 2015; 14: 389–401.
- [96] Shao MJ, Wang H, Dong L, Zhang HC, Lio X, Meng PS. Therapeutic Effect of Aloe-emodin Nanometer Liposome-mediated Photodynamic Therapy on Human Tongue Squamous Cell Carcinoma Tca-8113 Cells. *Journal of Oral Science Research*. 2018; 34: 948–951.
- [97] Abdissa N, Induli M, Fitzpatrick P, Alao JP, Sunnerhagen P, Landberg G, *et al.* Cytotoxic quinones from the roots of *Aloe dawei*. *Molecules (Basel, Switzerland)*. 2014; 19: 3264–3273.
- [98] Nigussie G, Melak H, Endale M. Traditional Medicinal Uses, Phytochemicals, and Pharmacological Activities of Genus *Rhamnus*: A review. *Journal of the Turkish Chemical Society Section A: Chemistry*. 2021; 8: 899–932.
- [99] Mai LP, Guéritte F, Dumontet V, Tri MV, Hill B, Thoison O, *et al.* Cytotoxicity of Rhamnosylanthraquinones and Rhamnosylanthrones from *Rhamnus nepalensis*. *Journal of Natural Products*. 2001; 64: 1162–1168.
- [100] Petit R, Izambart J, Guillou M, da Silva Almeida JRG, de Oliveira Junior RG, Sol V, *et al.* A Review of Phototoxic Plants, Their Phototoxic Metabolites, and Possible Developments as Photosensitizers. *Chemistry & Biodiversity*. 2024; 21: e202300494.
- [101] Muniyandi K, George B, Parimelazhagan T, Abrahamse H. Role of Photoactive Phytocompounds in Photodynamic Therapy of Cancer. *Molecules*. 2020; 25:4102.
- [102] Rajendran M. Quinones as photosensitizer for photodynamic therapy: ROS generation, mechanism and detection methods. *Photodiagnosis and Photodynamic Therapy*. 2016; 13: 175–187.

- [103] Maharjan PS, Bhattarai HK. Singlet Oxygen, Photodynamic Therapy, and Mechanisms of Cancer Cell Death. *Journal of oncology*. 2022; 2022: 7211485.
- [104] Okon E, Gawel-Bęben K, Jarzab A, Koch W, Kukula-Koch W, Wawruszak A. Therapeutic Potential of 1,8-Dihydroanthraquinone Derivatives for Breast Cancer. *International Journal of Molecular Sciences*. 2023; 24: 15789.
- [105] Khan AW, Farooq M, Haseeb M, Choi S. Role of Plant-Derived Active Constituents in Cancer Treatment and Their Mechanisms of Action. *Cells*. 2022; 11: 1326.
- [106] Malik MS, Alsantali RI, Jassas RS, Alsimaree AA, Syed R, Alsharif MA, *et al.* Journey of anthraquinones as anticancer agents - a systematic review of recent literature. *RSC Advances*. 2021; 11: 35806–35827.
- [107] Neganova M, Semakov A, Aleksandrova Y, Yandulova E, Pukhov S, Anikina L, *et al.* N-Alkylation of Anthracycline Antibiotics by Natural Sesquiterpene Lactones as a Way to Obtain Antitumor Agents with Reduced Side Effects. *Biomedicines*. 2021; 9: 547.
- [108] Min HY, Lee HY. Molecular targeted therapy for anticancer treatment. *Experimental & Molecular Medicine*. 2022; 54: 1670–1694.
- [109] Doytchinova I. Drug Design-Past, Present, Future. *Molecules (Basel, Switzerland)*. 2022; 27: 1496.
- [110] Gollnick K, Held S, Ma'rtire DO, Braslavsky SE. Hydroxyanthraquinones as sensitizers of singlet oxygen reactions: quantum yields of triplet formation and singlet oxygen generation in acetonitrile. *Journal of Photochemistry and Photobiology A: Chemistry*. 1992; 69: 155–165.
- [111] Montoya SCN, Comini LR, Sarmiento M, Becerra C, Albesa I, Argüello GA, *et al.* Natural anthraquinones probed as Type I and Type II photosensitizers: singlet oxygen and superoxide anion production. *Journal of Photochemistry and Photobiology B, Biology*. 2005; 78: 77–83.
- [112] Ortel B, Shea CR, Calzavara-Pinton P. Molecular mechanisms of photodynamic therapy. *Frontiers in Bioscience (Landmark Ed)*. 2009; 14: 4157–4172.
- [113] Aziz AK, Yousif TY. Characterization Studies on Mn(II), Co(II), Ni(II), Cu(II), Zn(II) and Cd(II) Complexes with New Ligands Derived from Anthraquinone Substance. *Iraqi Journal of Science*. 2022; 63: 2804–2813.
- [114] Mandal B, Mondal HK, Das S. In situ reactivity of electrochemically generated semiquinone on Emodin and its CuII/MnII complexes with pyrimidine based nucleic acid bases and calf thymus DNA: Insight into free radical induced cytotoxicity of anthracyclines. *Biochemical and Biophysical Research Communications*. 2019; 515: 505–509.
- [115] Tian W, Wang J, Zhong W, Huang H, Ji M, Yang T, *et al.* Synthesis, crystal structure and antitumor activity of trialkyltin (IV) 9,10-anthraquinone-2-carboxylate. *Journal of Organometallic Chemistry*. 2024; 1006: 122997.
- [116] Nowak-Perlak M, Ziółkowski P, Woźniak M. A promising natural anthraquinones mediated by photodynamic therapy for anticancer therapy. *Phytomedicine: International Journal of Phytotherapy and Phytomedicine*. 2023; 119: 155035.
- [117] Nowak-Perlak M, Bromke MA, Ziółkowski P, Woźniak M. The Comparison of the Efficiency of Emodin and Aloe-Emodin in Photodynamic Therapy. *International Journal of Molecular Sciences*. 2022; 23: 6276.
- [118] George BPA, Abrahamse H. A Review on Novel Breast Cancer Therapies: Photodynamic Therapy and Plant Derived Agent Induced Cell Death Mechanisms. *Anti-cancer Agents in Medicinal Chemistry*. 2016; 16: 793–801.
- [119] Kessel D. Apoptosis, Paraptosis and Autophagy: Death and Survival Pathways Associated with Photodynamic Therapy. *Photochemistry and Photobiology*. 2019; 95: 119–125.
- [120] Garg AD, Krysko DV, Verfaillie T, Kaczmarek A, Ferreira GB, Marysael T, *et al.* A novel pathway combining calreticulin exposure and ATP secretion in immunogenic cancer cell death. *The EMBO Journal*. 2012; 31: 1062–1079.
- [121] Sharma A, Sharma L, Nandy SK, Payal N, Yadav S, Vargas-De-La-Cruz C, *et al.* Molecular Aspects and Therapeutic Implications of Herbal Compounds Targeting Different Types of Cancer. *Molecules (Basel, Switzerland)*. 2023; 28: 750.
- [122] Dong X, Zeng Y, Zhang Z, Fu J, You L, He Y, *et al.* Hypericin-mediated photodynamic therapy for the treatment of cancer: a review. *The Journal of Pharmacy and Pharmacology*. 2021; 73: 425–436.
- [123] Gattuso H, Marazzi M, Dehez F, Monari A. Deciphering the photosensitization mechanisms of hypericin towards biological membranes. *Physical Chemistry Chemical Physics: PCCP*. 2017; 19: 23187–23193.
- [124] Kessel D. Hypericin Accumulation as a Determinant of PDT Efficacy. *Photochemistry and Photobiology*. 2020; 96: 1144–1147.
- [125] Kessel D. Photodynamic therapy: apoptosis, paraptosis and beyond. *Apoptosis: an International Journal on Programmed Cell Death*. 2020; 25: 611–615.
- [126] Kessel D. Death Pathways Associated with Photodynamic Therapy. *Photochemistry and Photobiology*. 2021; 97: 1101–1103.
- [127] Chua HM, Moshawih S, Goh HP, Ming LC, Kifli N. Insights into the computer-aided drug design and discovery based on anthraquinone scaffold for cancer treatment: A protocol for systematic review. *PLoS One*. 2023; 18: e0290948.
- [128] Siddamurthi S, Gutti G, Jana S, Kumar A, Singh SK. Anthraquinone: a promising scaffold for the discovery and development of therapeutic agents in cancer therapy. *Future Medicinal Chemistry*. 2020; 12: 1037–1069.
- [129] Fisher GR, Gutierrez PL, Oldcorne MA, Patterson LH. NAD(P)H (quinone acceptor) oxidoreductase (DT-diaphorase)-mediated two-electron reduction of anthraquinone-based antitumor agents and generation of hydroxyl radicals. *Biochemical Pharmacology*. 1992; 43: 575–585.
- [130] Liu Y, Liang Y, Jiang J, Qin Q, Wang L, Liu X. Design, synthesis and biological evaluation of 1,4-dihydroxyanthraquinone derivatives as anticancer agents. *Biorganic & Medicinal Chemistry Letters*. 2019; 29: 1120–1126.
- [131] Yu Y, Zhu L, Shi Y, Tong H, Kowah JAH, Wang L, *et al.* Novel anthraquinone photosensitizers: Synthesis, photoactivity, and 3D-QSAR studies. *Journal of Molecular Structure*. 2023; 1292: 136107–136119.
- [132] Sulaiman C, George BP, Balachandran I, Abrahamse H. Photoactive Herbal Compounds: A Green Approach to Photodynamic Therapy. *Molecules (Basel, Switzerland)*. 2022; 27: 5084.
- [133] Woźniak M, Nowak-Perlak M. Hypericin-Based Photodynamic Therapy Displays Higher Selectivity and Phototoxicity towards Melanoma and Squamous Cell Cancer Compared to Normal Keratinocytes In Vitro. *International Journal of Molecular Sciences*. 2023; 24: 16897.